



Research Update: Pollination Speed Talks

December 8, 2015



Speakers

Bob Curtis, Almond Board (Moderator)

Steve Sheppard, Washington State University

Jody Johnson, Cullaborate

Neal Williams, University of California, Davis

Ellen Topitzhofer, Oregon State University

Carolyn Breece, Oregon State University

Quinn McFrederick, University of California,
Riverside

Fabiana Ahumada, Ag Science Consulting

Troy Anderson, Virginia Tech





**Bob Curtis,
Almond Board**



**Steve Sheppard,
Washington State University**

Honey Bee Stock Improvement

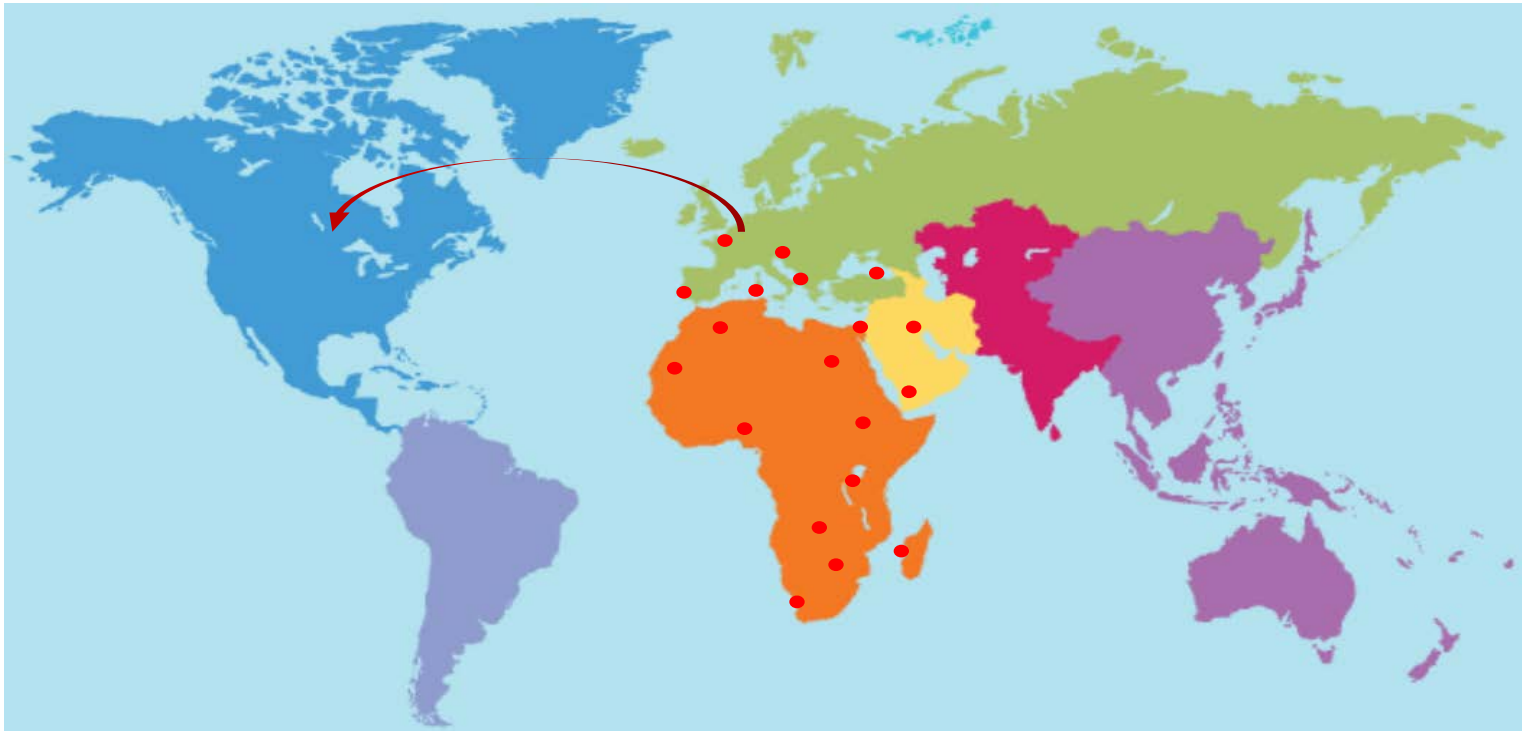
S. Sheppard, S. Cobey, B. Hopkins
Department of Entomology
Washington State University
Pullman WA



Almonds and Honey Bees – Both have overseas origins !



A subset of more than 24 Old World honey bee subspecies form the basis for all current US populations



Only three introduced subspecies were maintained by US beekeepers

Subsp.	Origin	Arrival
<i>mellifera</i>	Europe	1600's
<i>ligustica</i>	Europe	1859
<i>lamarckii</i>	Africa	1866
<i>carnica</i>	Europe	1877
<i>cypria</i>	Middle East	1880
<i>syriaca</i>	Middle East	1880
<i>caucasica</i>	Europe	1880-1882
<i>intermissa</i>	Africa	1891
<i>scutellata</i>	Africa	1990

1922 Honey Bee Act

Restricted further importation of honey bees into the U.S. in an attempt to keep out tracheal mites

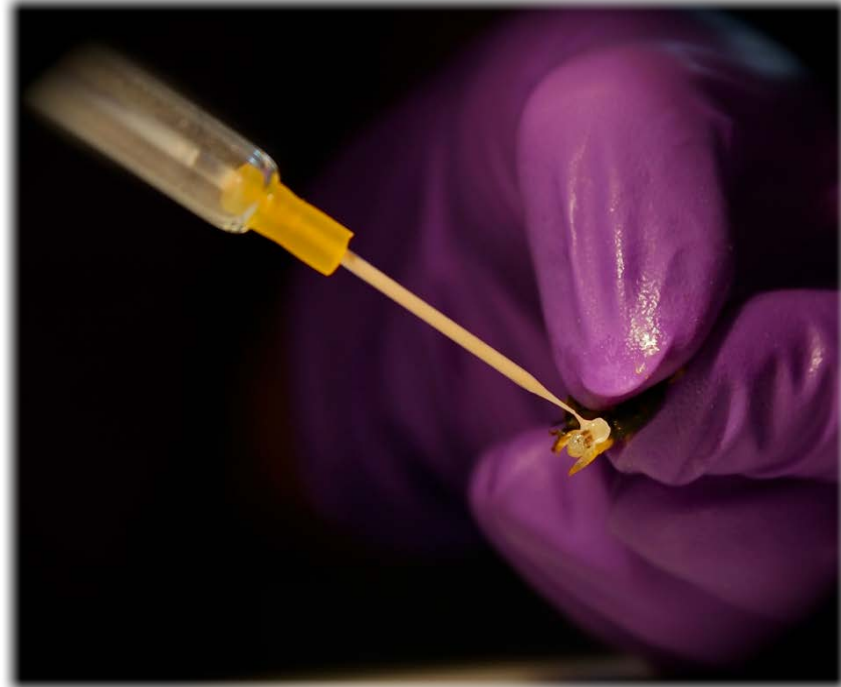


Honey bee breeding and stock improvement program - progress

- Importation of additional genetic diversity – Old World source populations
- Cryogenetic methods for honey bee semen
- Establishment of a honey bee germplasm repository
- Incorporation and distribution of novel genetics to honey bee queen producers
- Reestablishment of Caucasian honey bees
- Introduction of a new subspecies adapted to cold weather in 2015 – *A. m pomonella*



Semen collection, cryopreservation, instrumental insemination



2015 – Tien Shan Mountains, Kazakhstan
A. m. pomonella









Questions?



Jody Johnson, Cullaborate



The Effect of Application Time on Fungicide Exposure to Honey bees in Almonds

Pettis, J., Bluher, S., Johnson, J., Wardell, G.
Dec. 8, 2015



Why study fungicides in honey bees?

Synergistic relationships of insecticides with fungicides (Johnson et al. 2013)

Fungicide loads in bee-collected pollen correlate with higher loads of *Nosema* (Pettis et al. 2013)



Iprodione (a carboximide)
inhibits DNA and RNA synthesis,
cell division and cell metabolism.

Rovral 4F was applied by air blast
ground rig at a uniform rate.

Two applications:

First: Site 1 at 6pm on Day 1

Second: Site 2 at 11am on Day 3.

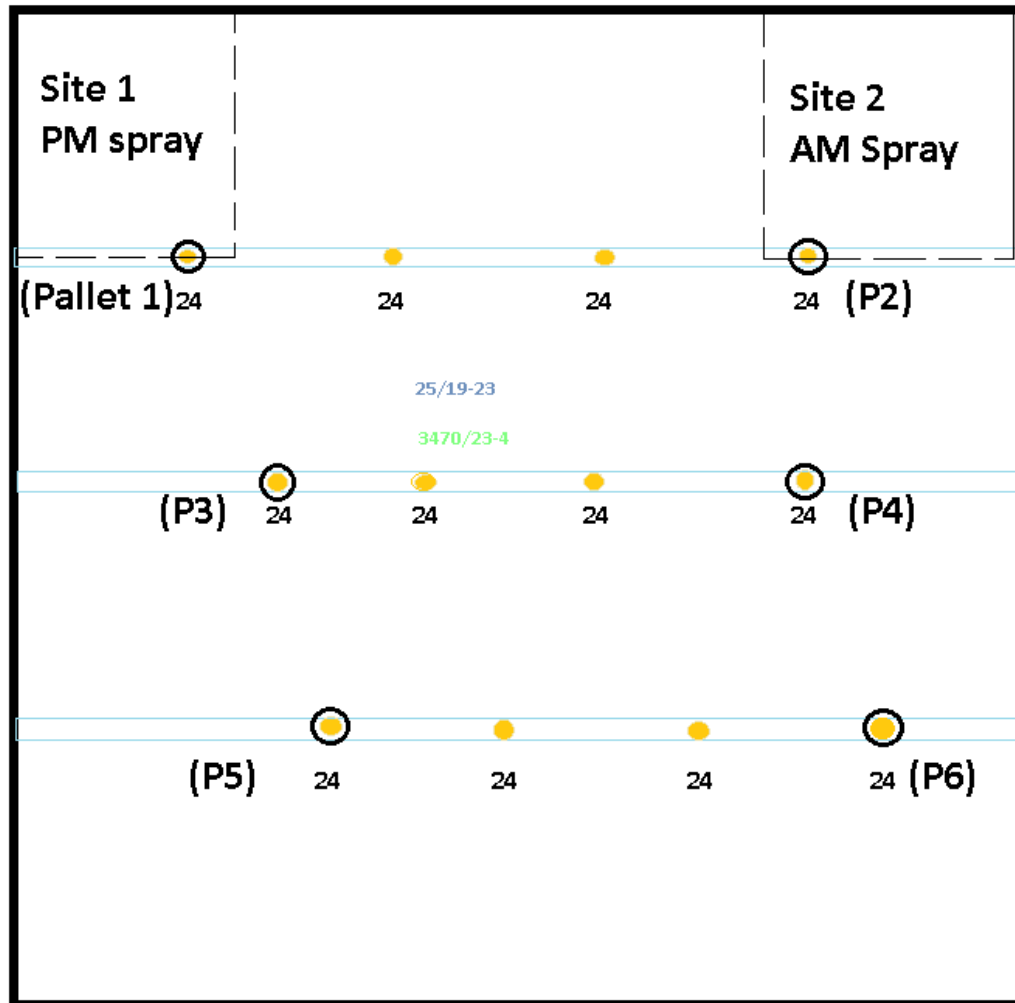
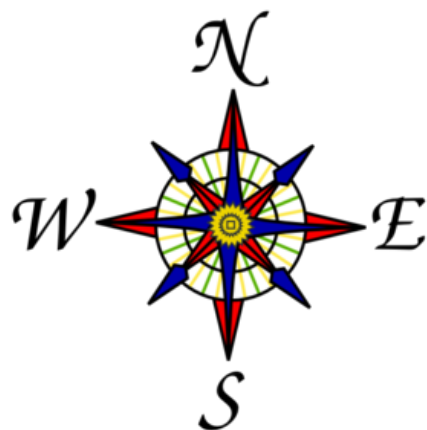


Objective

To determine if spraying fungicide at different times of day (AM vs. PM) leads to differences in the exposure levels to foraging honey bees and bee-collected pollen



Orchard map





Foraging Counts

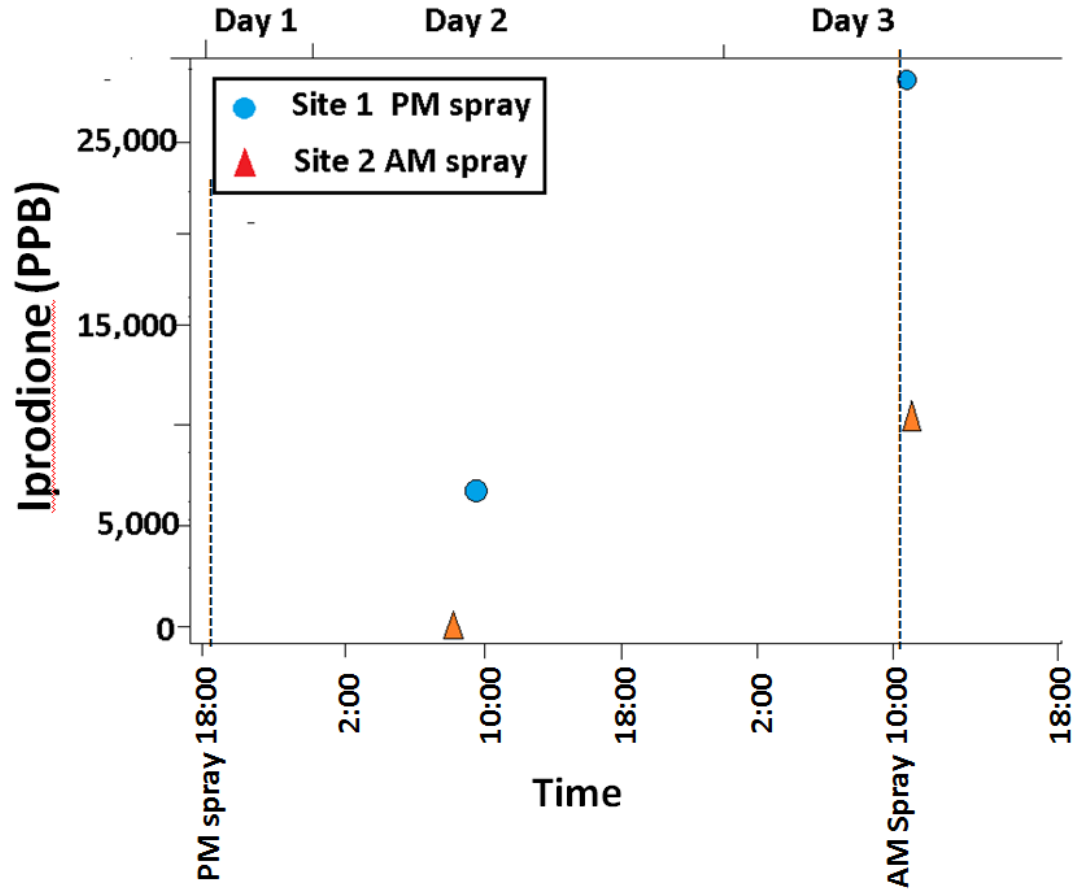
	Day 1	Day 2	Day 3
Pollen Foragers	39	27	13
Nectar Foragers	114	105	219
Foragers in Trees	6	3.6	2.8

Table 1: Mean foraging counts across the three days of study. Pollen and nectar foragers counted returning to hive entrance during three-minute interval, differentiated by presence/absence of corbicular load.

Iprodione levels (ppb) were monitored in anther pollen after spray events.



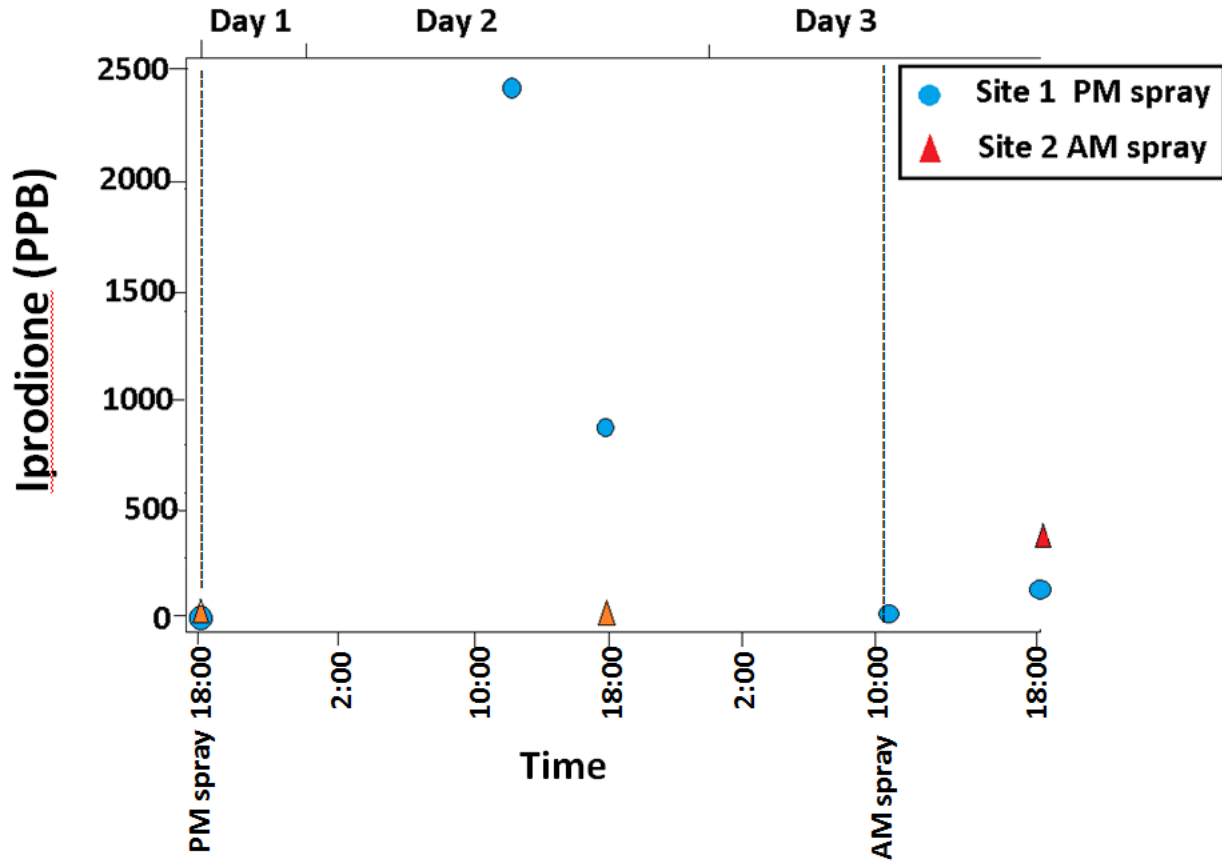
Anther pollen



Iprodione levels (ppb) were monitored in forager-collected pollen for the three day study



Forager-collected pollen



Conclusions

- Anther pollen contained higher loads of iprodione following AM spray
→ Greater potential for exposure during foraging hours following AM spray vs PM spray. High iprodione in PM site area after AM spray may be due to drift.
- Actual exposure to iprodione in hives was lower following AM spray vs PM spray → decreased foraging activity or reduction of visitation to almonds due to diminishing availability of bloom.
- This study occurred during last week of bloom. Study should be undertaken during peak or consistent bloom.



Thank you

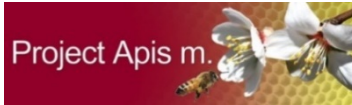
**Almond Board of California for funding the study
Paramount Farms for hosting the study**



**Neal Williams,
University of California, Davis**

Fungicide Residual Effects on Fertilization through Stigma-Receptivity, Pollen Germination, and Tube Growth

Neal M. Williams
University of California, Davis



Fungicide timing- defining the issue

- Cool moist weather
- Ideal conditions for fungal pathogens
- Infection through blossoms

- Fungicides applications following rains are an integral part of best management for almond

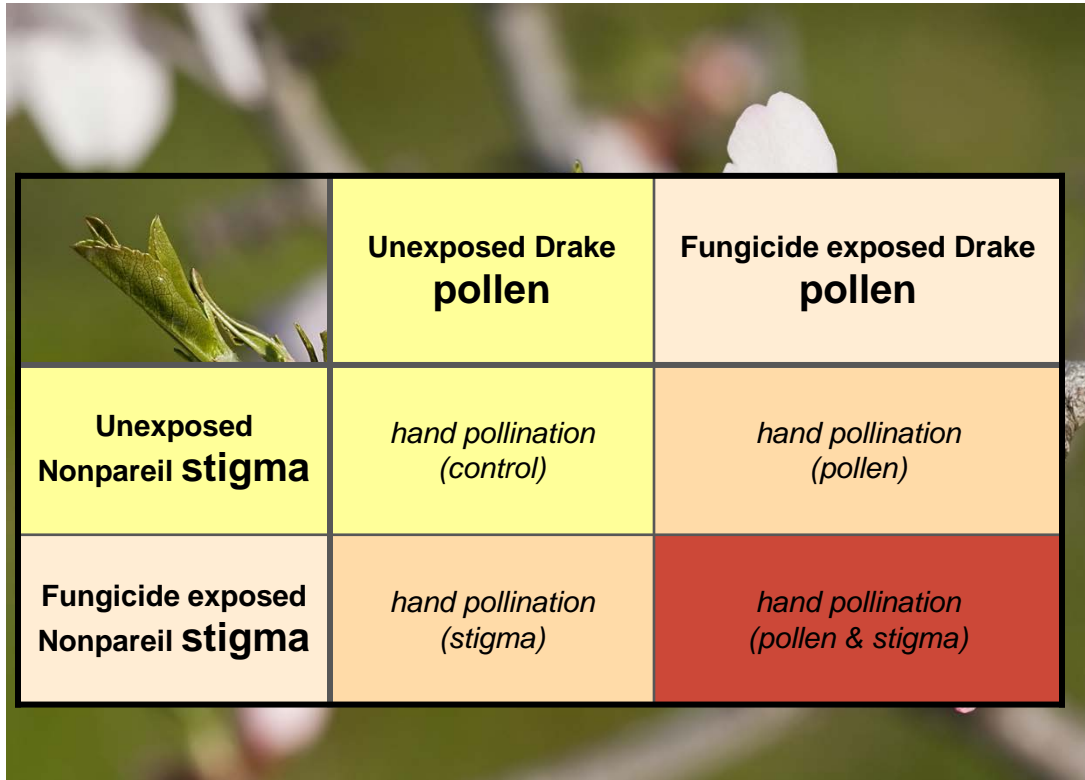
- Optimal timing for spray application?
- Differences in impact of fungicides
- on POST POLLINATION aspects



Flower stages and fungicide exposure



Study Design



	Unexposed Drake pollen	Fungicide exposed Drake pollen
Unexposed Nonpareil stigma	<i>hand pollination (control)</i>	<i>hand pollination (pollen)</i>
Fungicide exposed Nonpareil stigma	<i>hand pollination (stigma)</i>	<i>hand pollination (pollen & stigma)</i>

Study Design

Two New Fungicide Classes

FRAC 3 - demethylation inhibitor

FRAC 7 - succinate
dehydrogenase inhibitor

New 2016

FRAC 9 - methionine biosynthesis

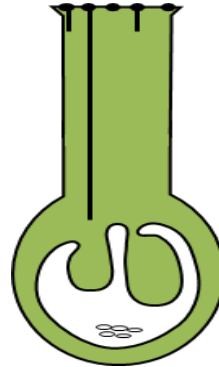
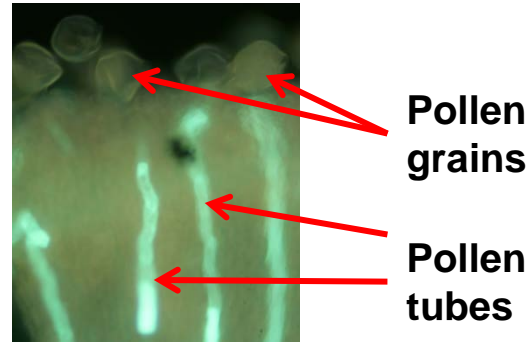
FRAC 11 - quinone outside inhibitor



Study Methods

Assess differences in ovule fertilization of intact flowers from the different treatments

1. Timing (open flower versus in bud)
2. Before and after pollination
3. Pathways of effect: Pollen vs Stigma / style

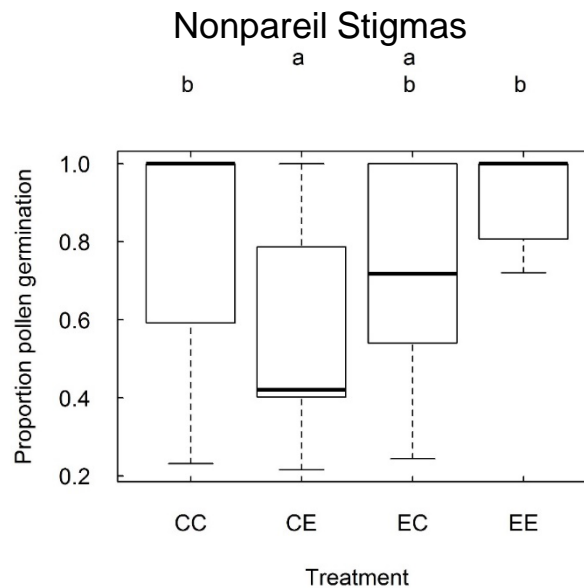
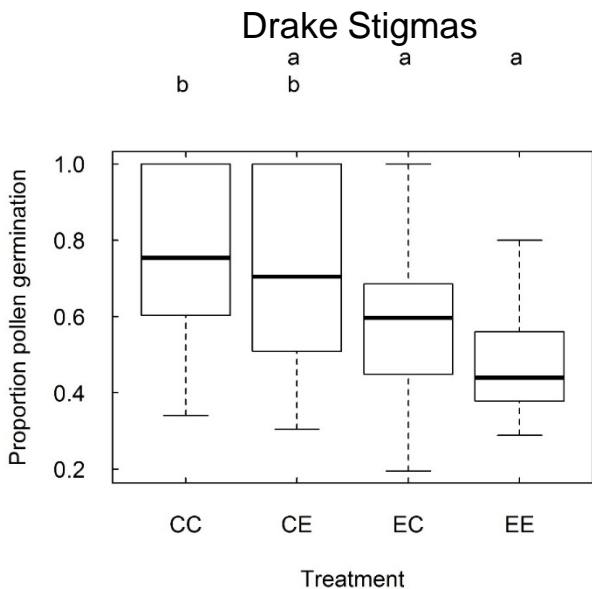


Results (2013)

Stigma variety	Chemical	Exposure	Pollen germination	Pollen tube development	No. pollen tubes ovary
Drake	FRAC 3	bud	0.210	†0.166	0.337
Drake	FRAC 3	flower	0.321	0.878	0.506
Drake	FRAC 7	bud	0.003	0.655	0.982
Drake	FRAC 7	flower	0.994	†0.969	0.952
Nonpareil	FRAC 3	<u>bud</u>	0.004	†0.176	0.115
Nonpareil	FRAC 3	flower	0.816	0.517	0.921
Nonpareil	FRAC 7	<u>bud</u>	0.066	†0.089	0.224
Nonpareil	FRAC 7	flower	0.861	0.510	0.383

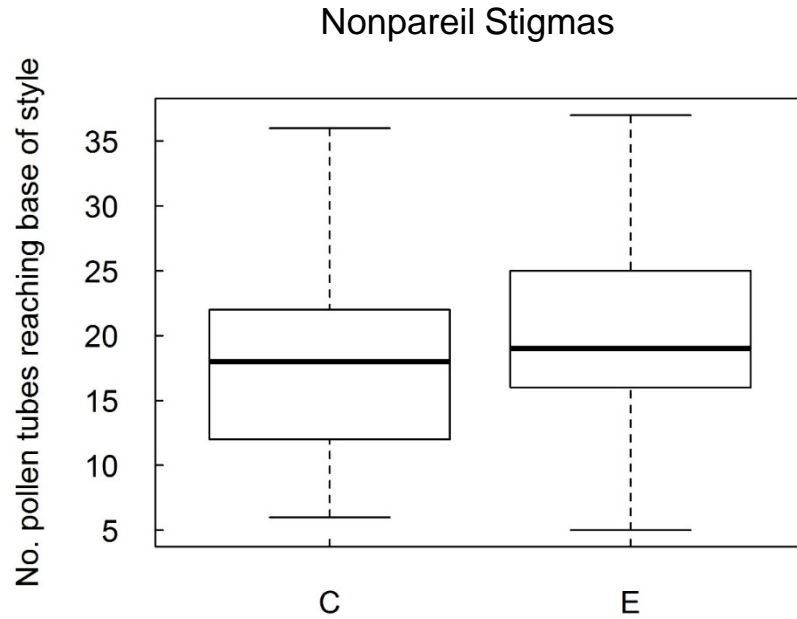
Results- Spray precedes pollination

Pollen germination



CC= control, CE= control stigma exposed pollen, EC= exposed stigma control pollen, EE= exposed stigma exposed pollen

Results- Post-pollination sprays



C= control , E= exposed stigma

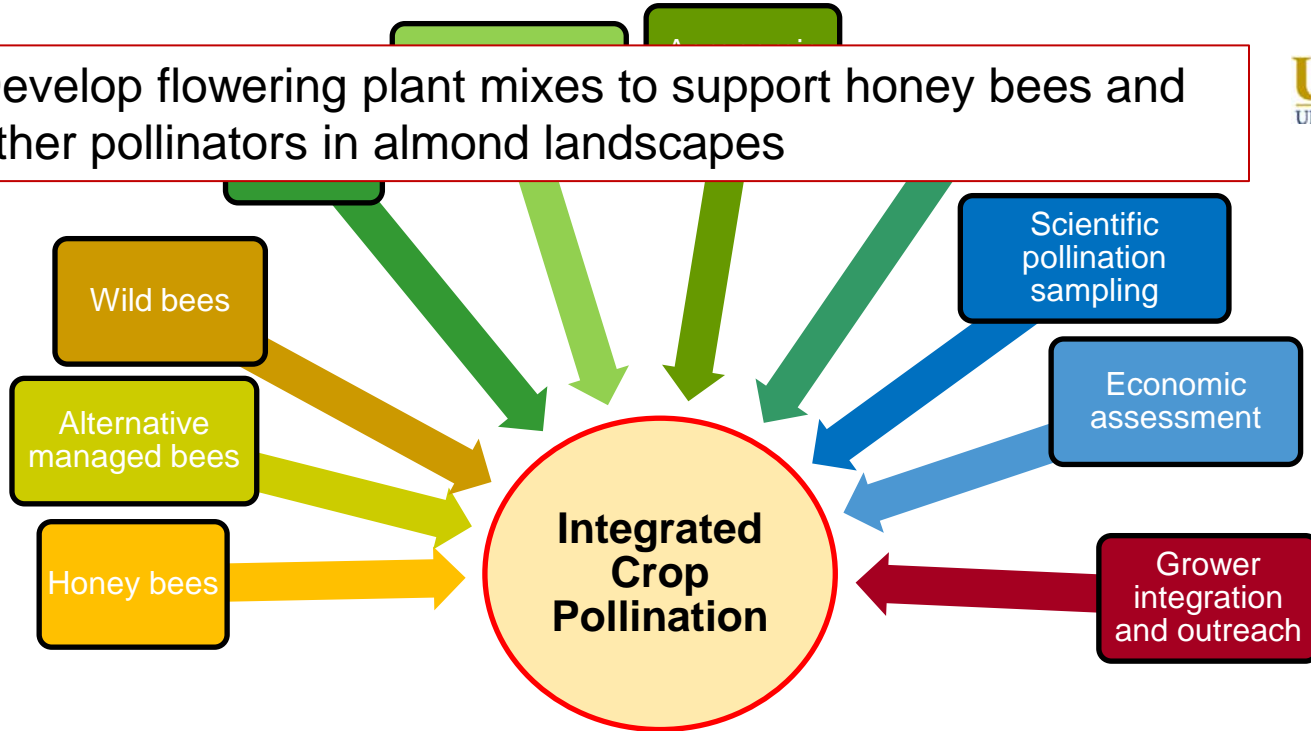
Summary

- Impacts of FRAC 3 and FRAC 7 fungicides are modest and inconsistent
- Decrease in pollen germination, does not persist through fertilization
- Streamline method for fungicide testing that could be easily and more widely applied to new sprays
- Additional FRAC groups 9, 11 will be tested

Forage for honeybees: Integrated Crop Pollination



Develop flowering plant mixes to support honey bees and other pollinators in almond landscapes



Highlights

- **Mustard and wildflower** mixes provided the **most bloom** and wildflower flowering persisted longer after almond flowering
- **Mustard and wildflower mix attracted** the **most honeybees**
- **Wildflower** mix, then mustard **attracted** the **most wild bees**
- Mixes **did not** attract honey bees away from the orchard flowers



A close-up photograph of several green almonds on a branch, with vibrant green leaves. The background is softly blurred, showing more of the tree and a hint of a person in the distance.

**Ellen Topitzhofer, Oregon
State University**



Tech Transfer Teams for Commercial Beekeeping: Pacific Northwest Team

Ellen Topitzhofer

PI: Ramesh Sagili

Oregon State University

The Bee Informed Partnership



Using beekeepers' real world experience to solve
beekeepers' real world problems

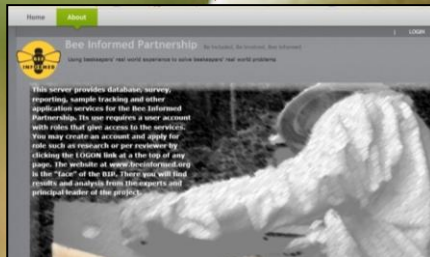
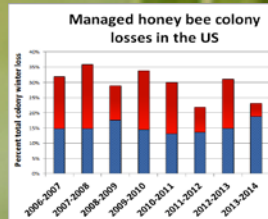


United States Department of Agriculture
National Institute of Food and Agriculture



On-the-ground
testing

Surveys



Database



National historic and
on-going diagnostic data



4 visitations for our commercial beekeepers

Treatment

Treatment

1

Feb/
March



2

April/
June

3

July/
August



4

Sept/
Oct

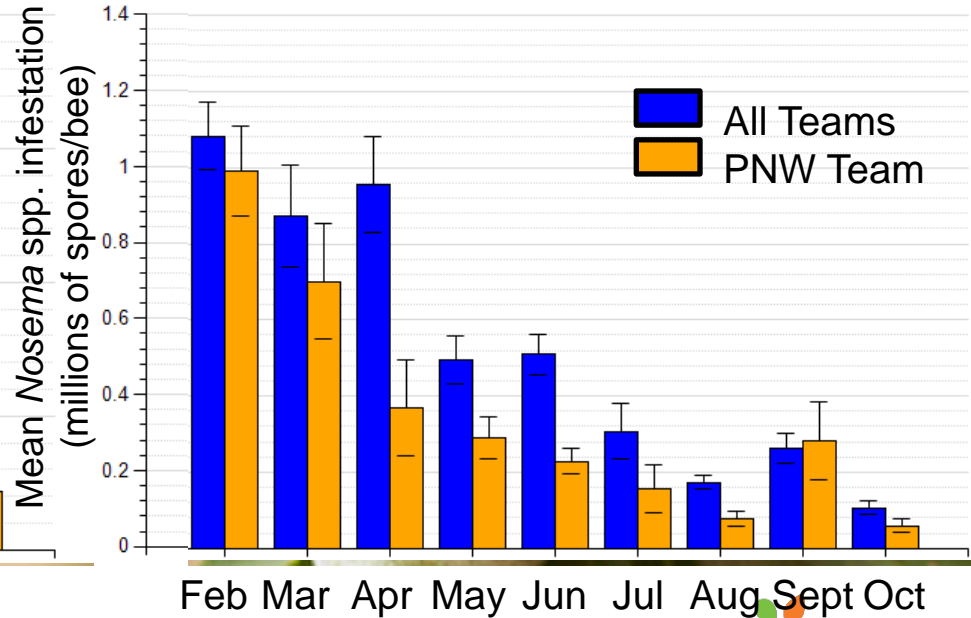
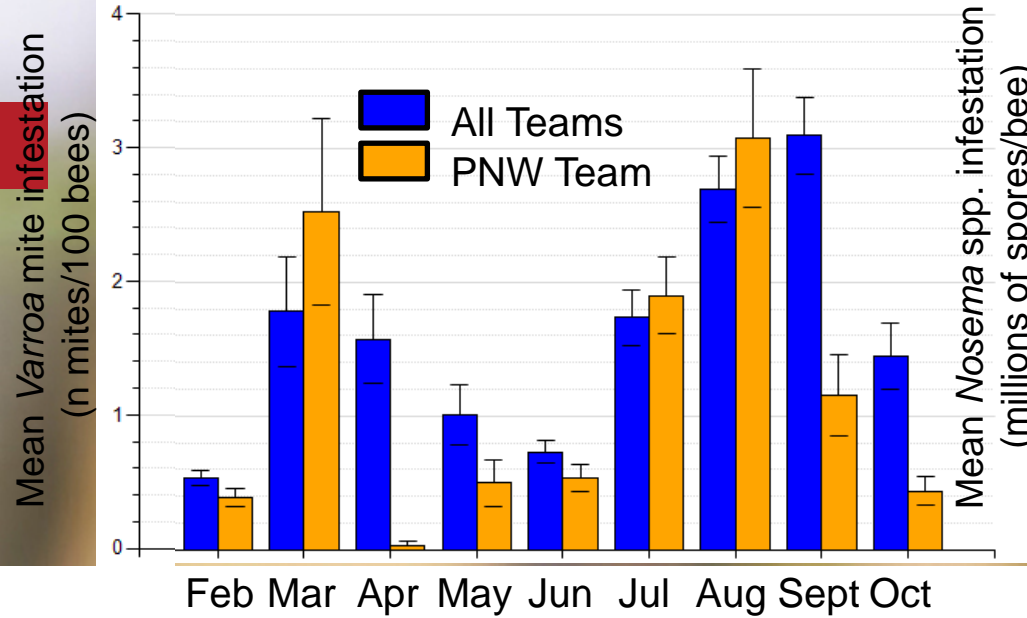
Sample Types

- Varroa/Nosema: % Varroa mite infestation and Nosema spp. spore count
- Viruses: quantify levels of 7 viruses (NC State)
- Protein: head protein content (OSU)
- Queen quality: sperm viability and count (NC State)
- Disease (ABF/EFB): presence/absence (USDA-Beltsville)
- Pesticides: quantify and report as PPB (USDA-Gastonia)



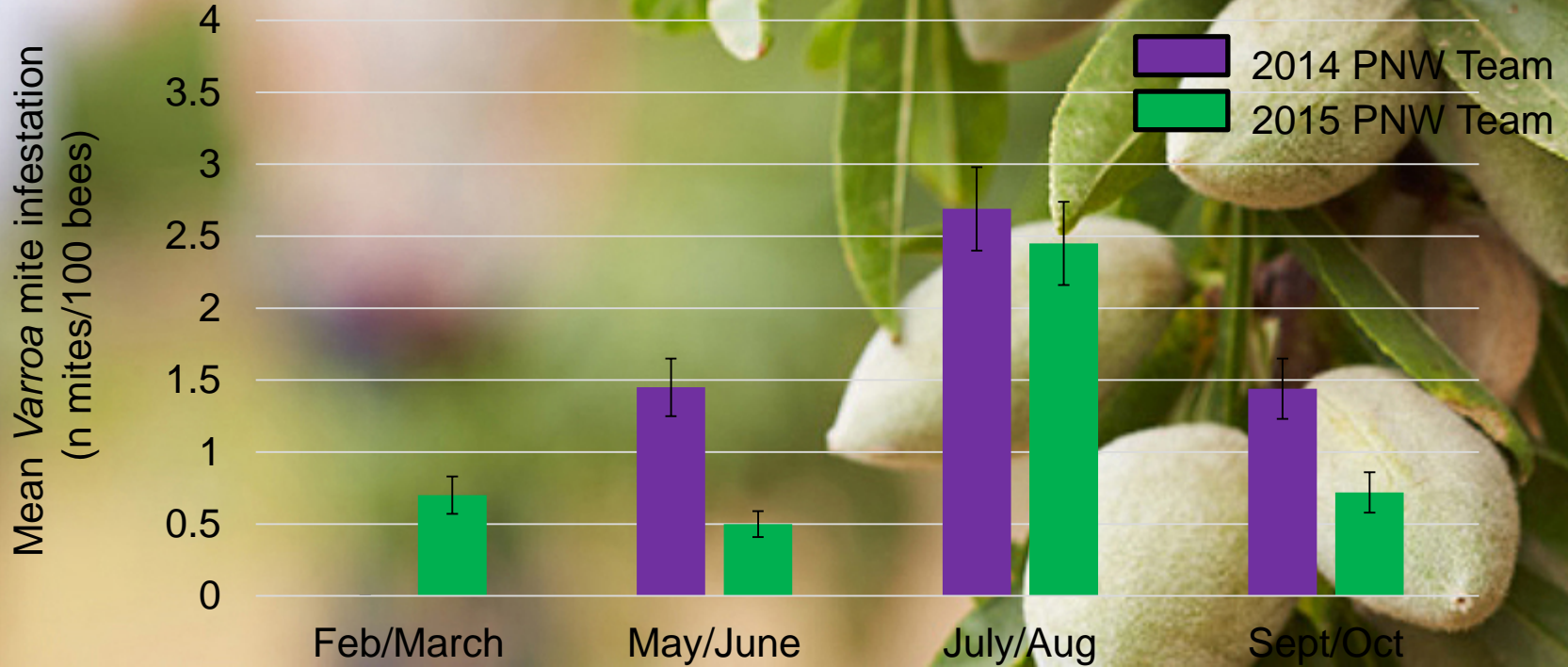
2015 Varroa sample levels

2015 Nosema levels



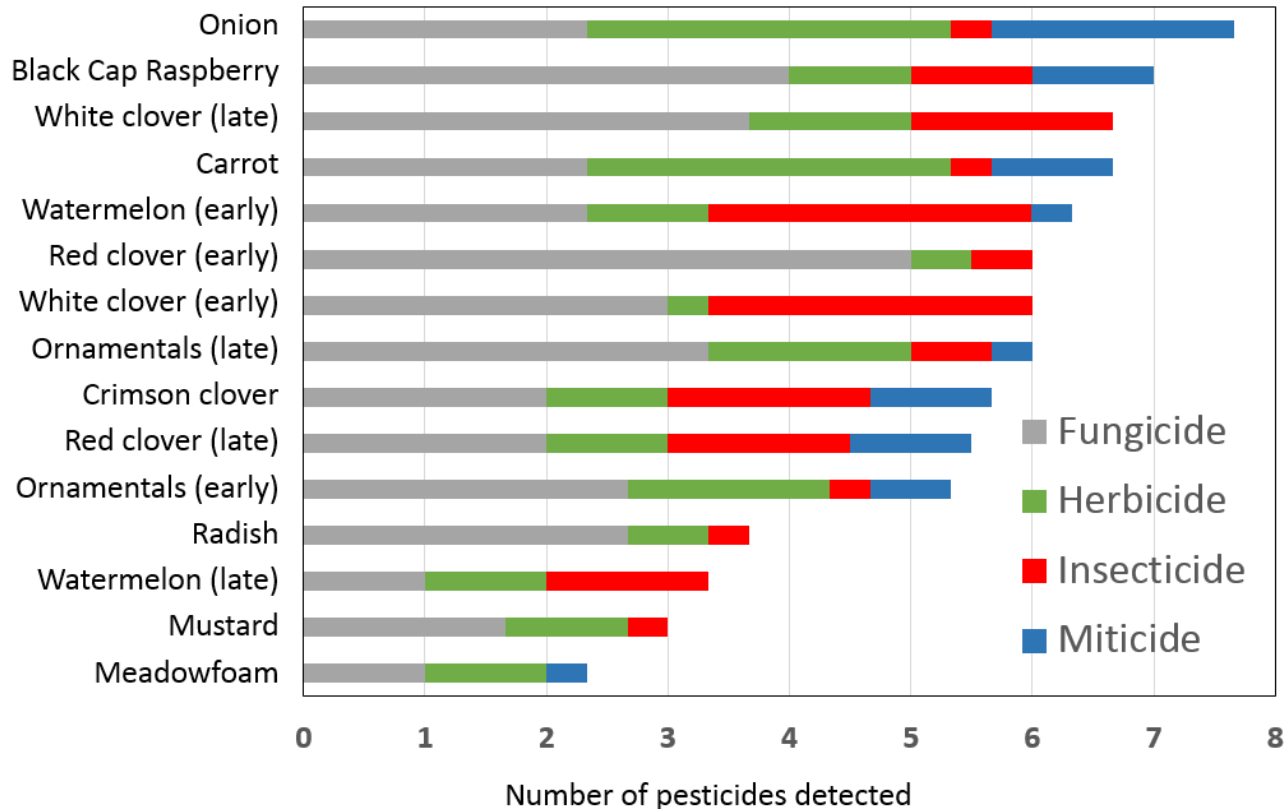
Error bars indicate 95% CI

2014 vs. 2015 Varroa levels

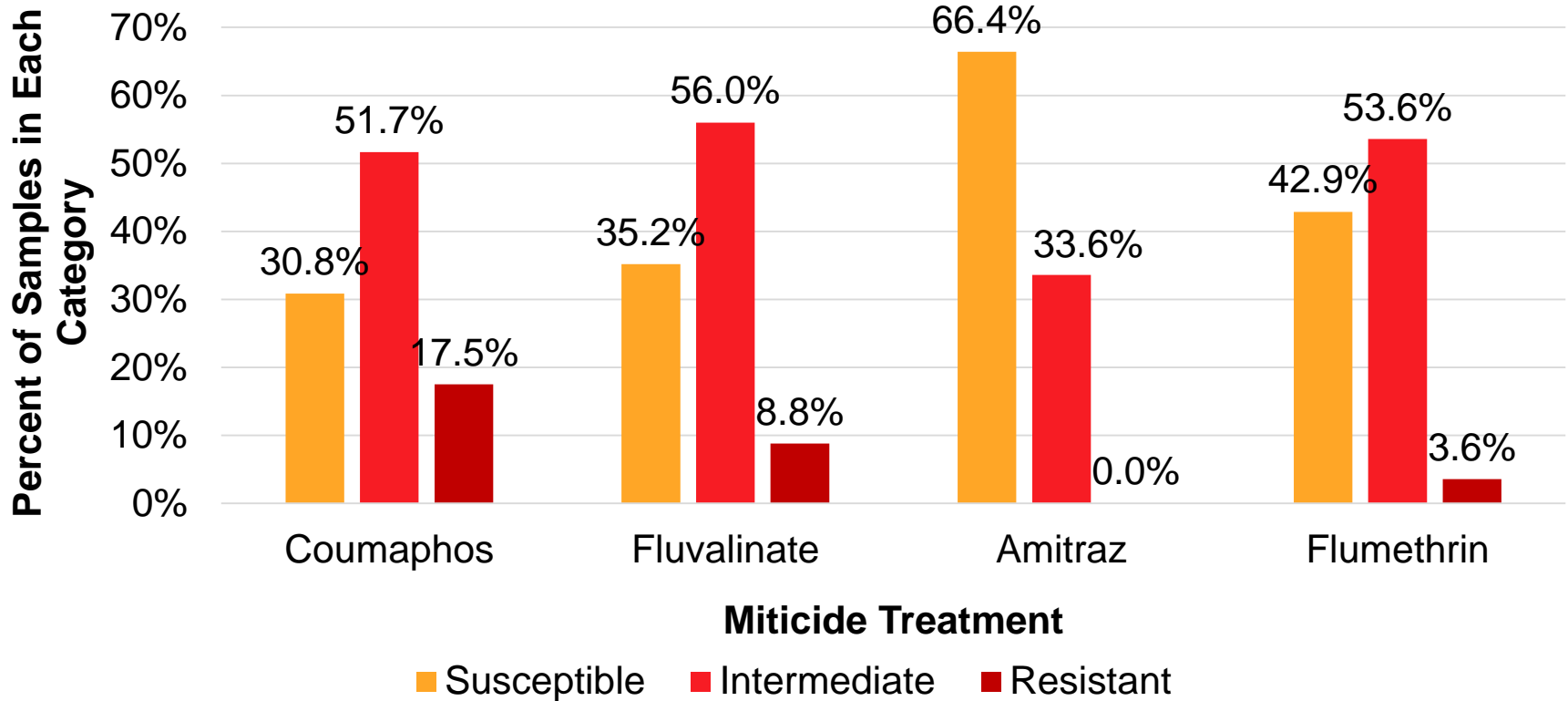


Error bars indicate 95% CI

Total pesticides detected per field



National Breakdown between Resistant, Intermediate and Susceptible Varroa (2012-2014)



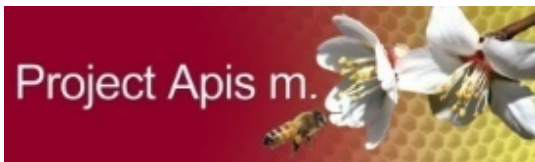


United States Department of Agriculture
National Institute of Food and Agriculture

Thank You!



AlmondBoard.com



Carolyn Breece, Oregon State University



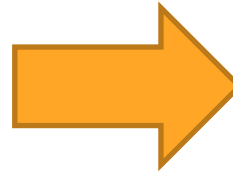


Assessing the value of supplemental forage for honey bees during almond pollination

Ramesh Sagili and Carolyn Breece
Oregon State University



Planting supplemental forage for honey bees

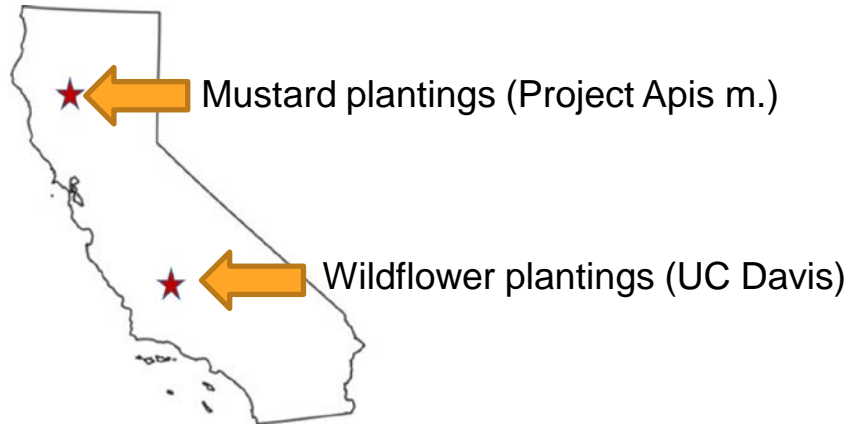




Objective: To evaluate the effects of supplemental forage prior to and after almond bloom on honey bee nutrition, colony growth, immune system and survival.



Methods



Methods

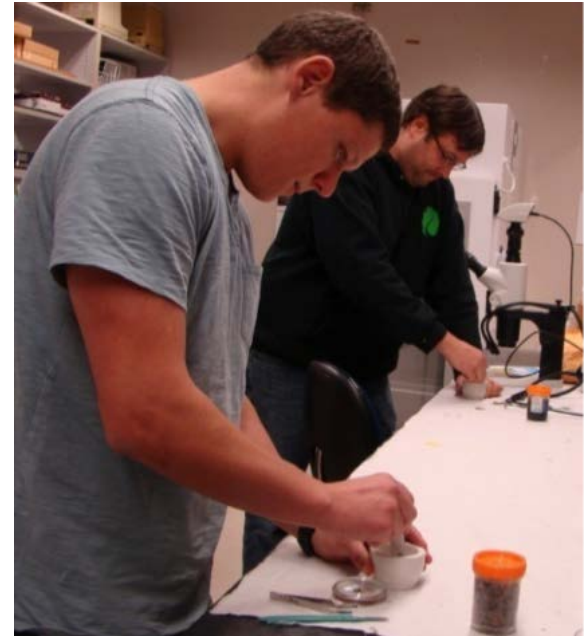
- Colony evaluations
- Pollen traps
- Honey bee samples
- Lab analysis



Results

We analyzed bee samples for

- varroa mites
- nosema
- protein content (measure of nutrition)
- immune system enzymes



Results were highly variable.

Results: Pollen identification and proportion

Wildflower	Site	Pollen	23 Feb	27 Feb	6 Mar	10 Mar
Far	MA	Almond	94	76	6	
		Wildflowe	3	9	42	
		mustard	0	7	6	
		Unknown	3	5	20	
		Weed	0	3	31	
Near	MW	Almond	72	58	0	0
		Wildflowe	7	15	8	23
		mustard	16	42	23	57
		Unknown	4	6	6	2
		Weed	1	9	66	18

Mustard			12 Feb	21 Feb	12 Mar
Far	HA	Almond	97	96	76
		mustard	0	0	14
		Unknown	3	4	86
		Weed	0	0	0
	KA	Almond	91	50	0
		mustard	1	48	91
		Unknown	8	3	12
		Weed	0	0	1
Near	KM	Almond	91	31	0
		mustard	0	66	64
		Unknown	10	8	36
		Weed	0	0	0
	PM	Almond	93	31	1
		mustard	3	73	83
		Unknown	4	6	77
		Weed	0	0	0

Tables courtesy of Kimiora Ward and Neal Williams, UC Davis

Thank you

Our collaborators:

- Dr. Neal Williams and Kimiora Ward, U.C. Davis
- Project Apis m.
- Wonderful Farms
- Beekeepers from California and Oregon
- Almond growers

We thank Almond Board of California for providing funds for this project.





**Quinn McFrederick,
University of California, Riverside**

The influence of cover crop forage on honey bee nutrition and gut microbes, and on colony growth and activity

Quinn McFrederick¹, William Meikle², Mark Carroll²

1. UC Riverside Department of Entomology
2. USDA Carl Hayden Bee Research Center



Objectives and methods

Objectives: Determine effects of supplemental rapini forage on honey bee:

- 1) Nutrition, health, and queen quality
- 2) Brood production
- 3) Gut microbiome
- 4) Interactions between all these factors

- Methods
 - 40 colonies
 - 2 forage and 2 non-forage plots in AZ
 - mid January
 - Moved to Almonds
 - mid February

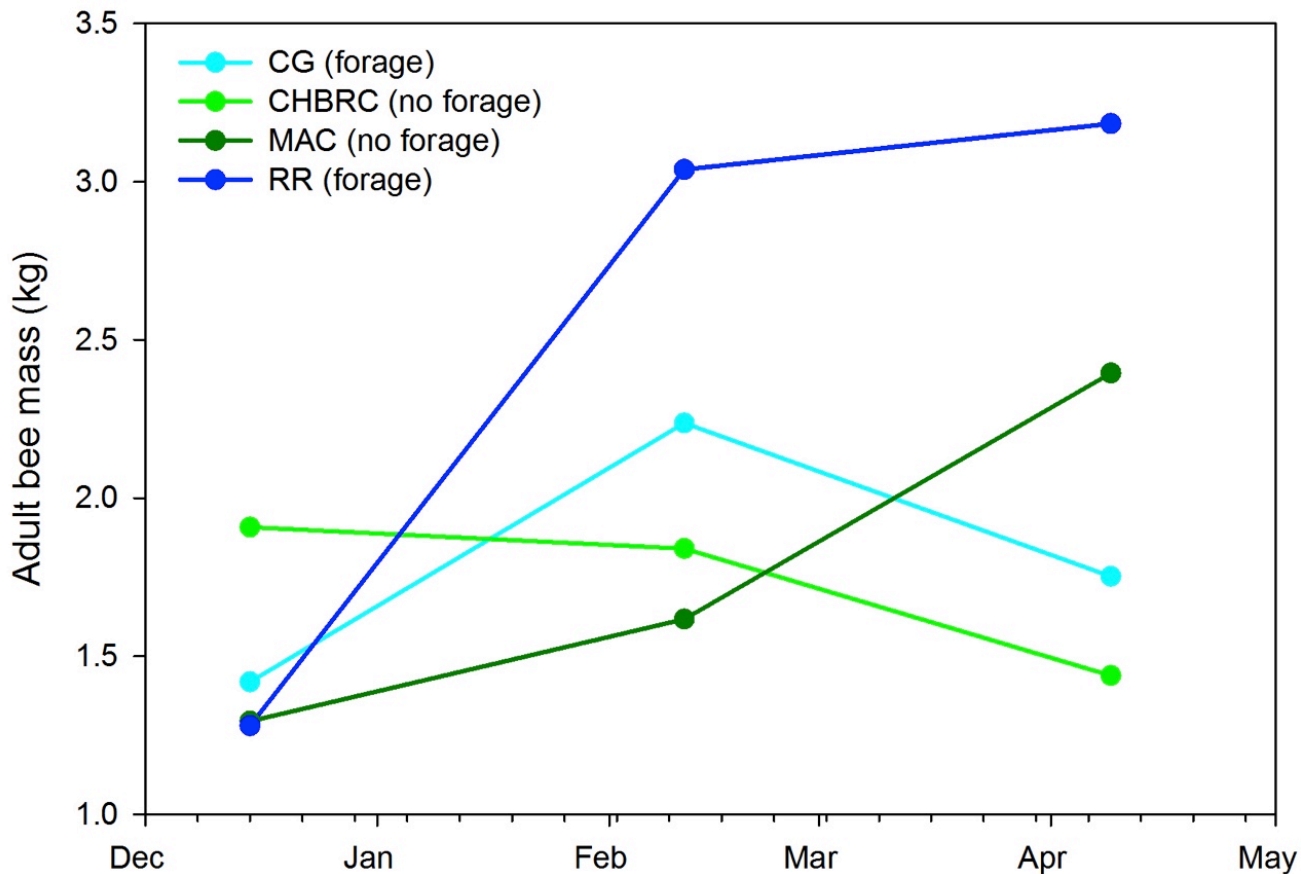


Results

Average adult bee mass

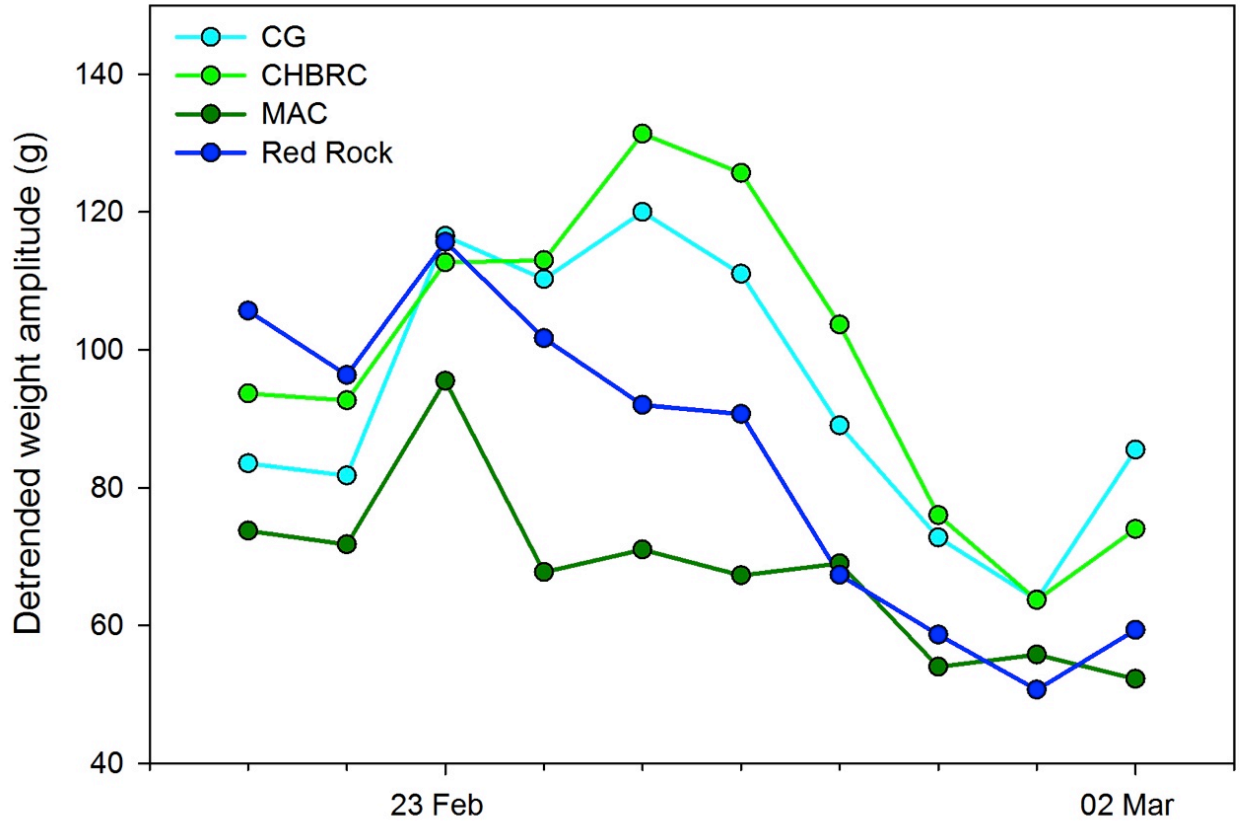


Adult bee mass 2014-15



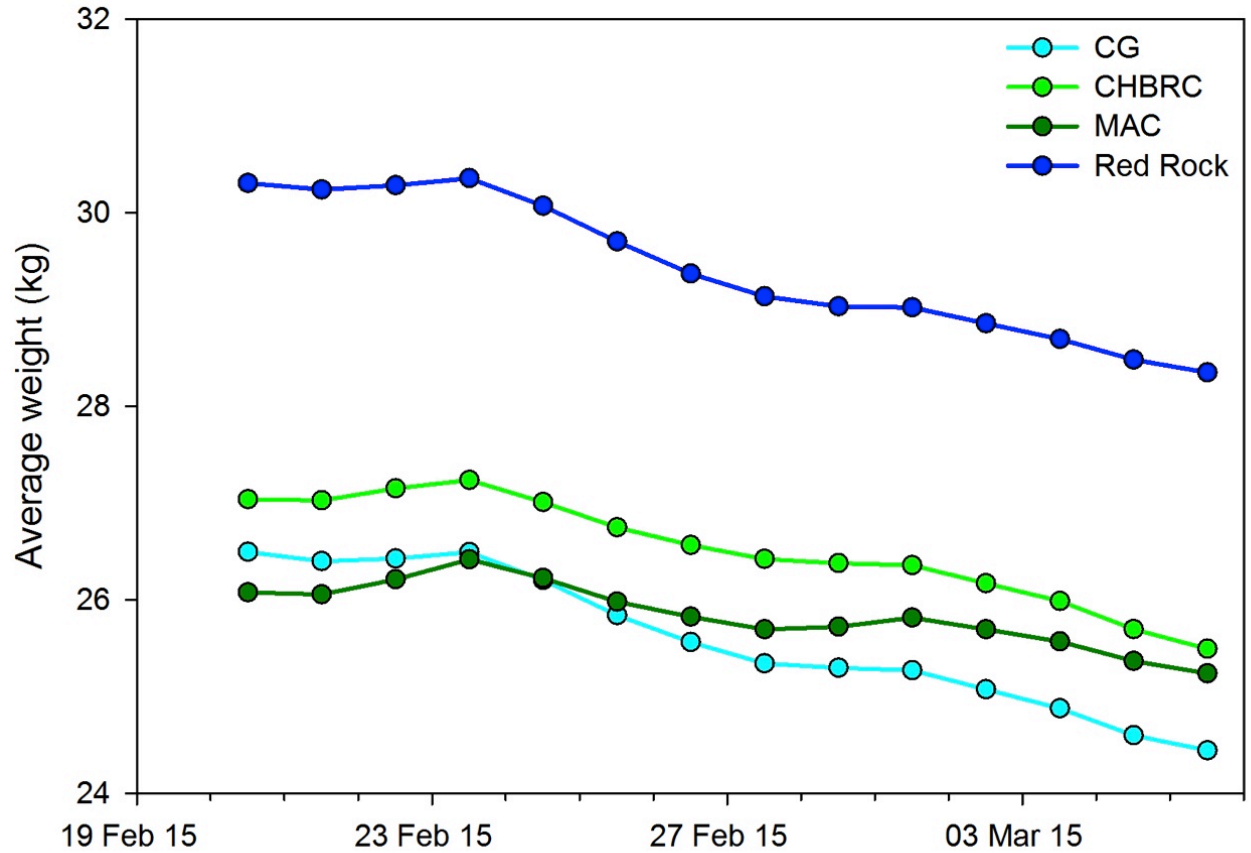
Results

Daily colony weight variation during almond pollination



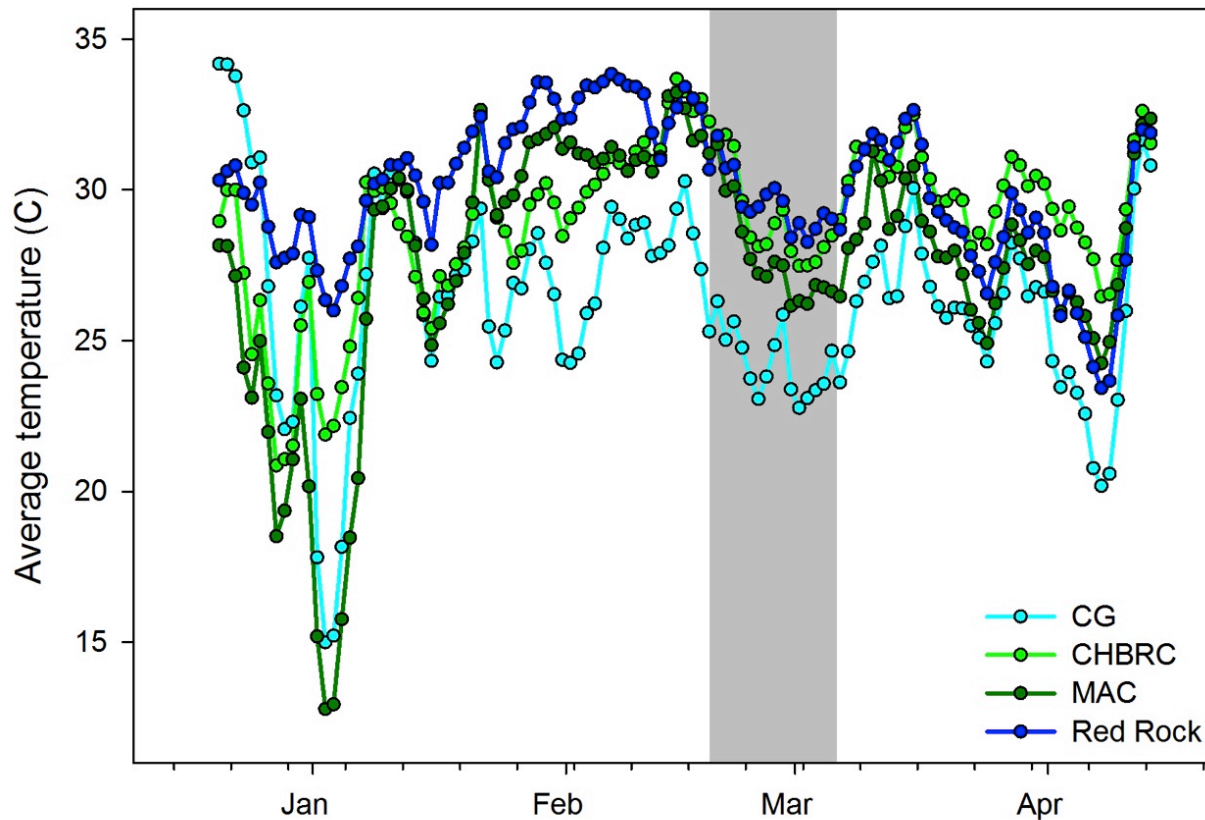
Results

Average weight during almond pollination



Results

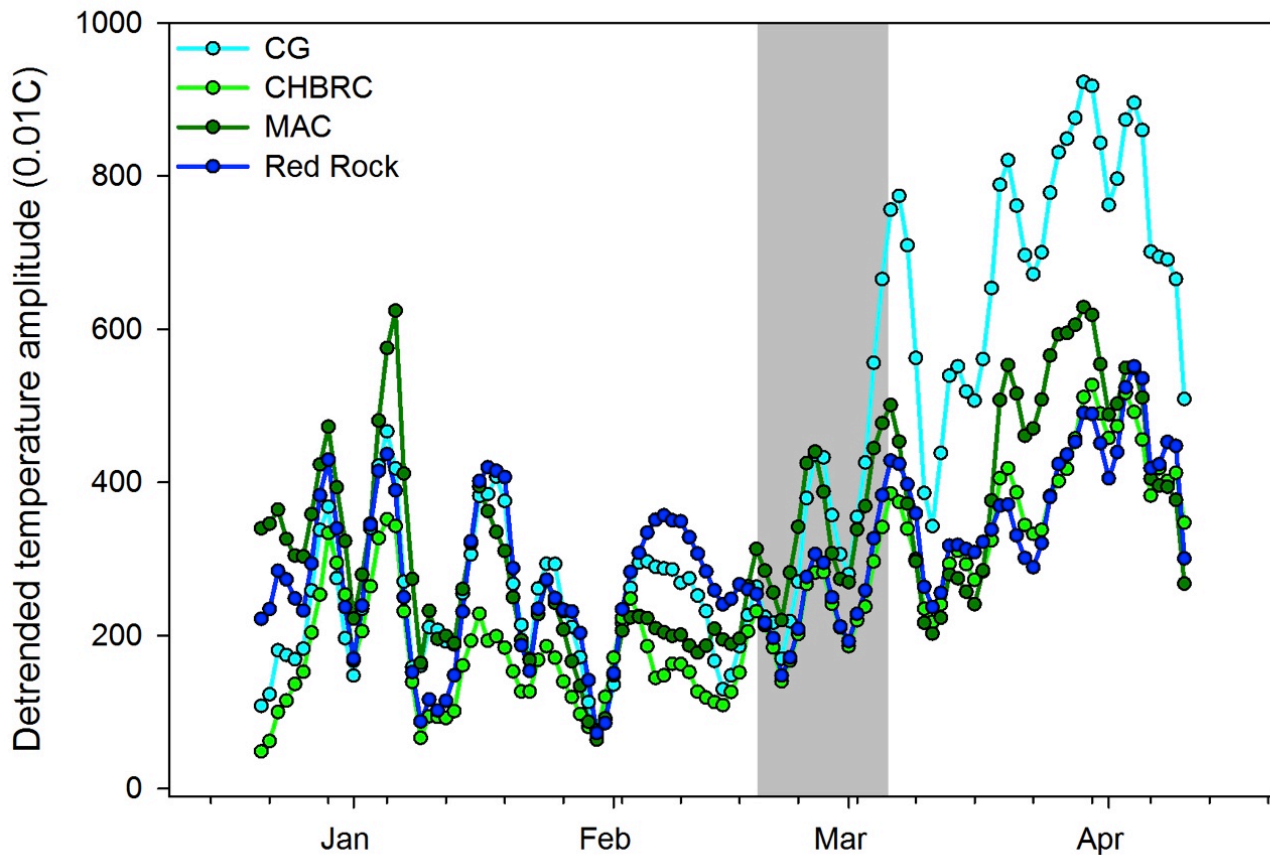
Average temperature inside the colony



Results

Average daily temperature variation

- Lower numbers indicates brood rearing



Conclusions

- Forage treatment did not increase weight
 - Caveats
 - Non-forage sites had pollen coming into hives
 - Forage treatment not as long as planned
- Site matters
 - Red Rock colonies had more foragers and greater weight
- Nutrition and gut microbe work ongoing
 - 1,100 honey bees dissected
 - ~200 samples awaiting DNA sequencing
 - Queen quality



Thanks to Milagra Weiss, Nick Brown, Jason Rothman, and Wonderful Farms for logistical support, and to you for your support.



**Fabiana Ahumada,
Ag Science Consulting**

Implementing an Integrated Pest Management for *Varroa*

Fabiana Ahumada
AgScience Consulting



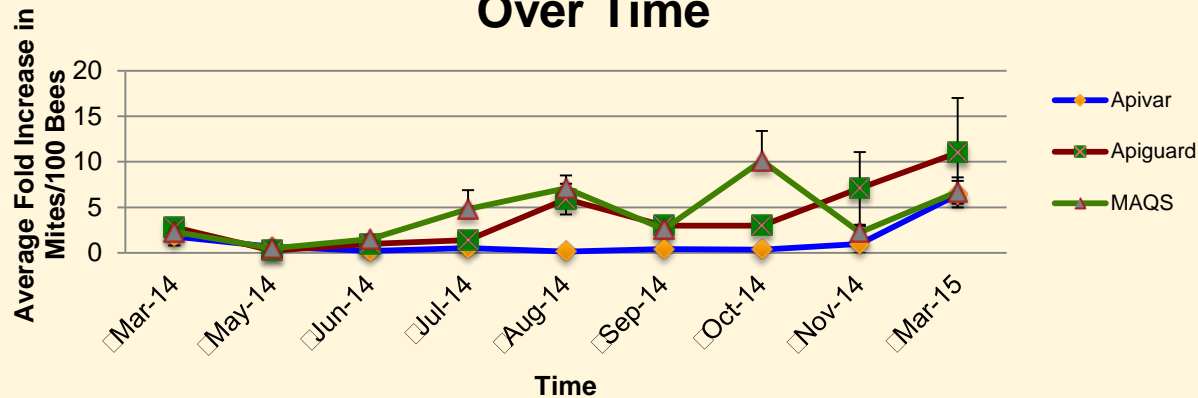
Project Overview

Implement an Integrated Pest Management program for Varroa control

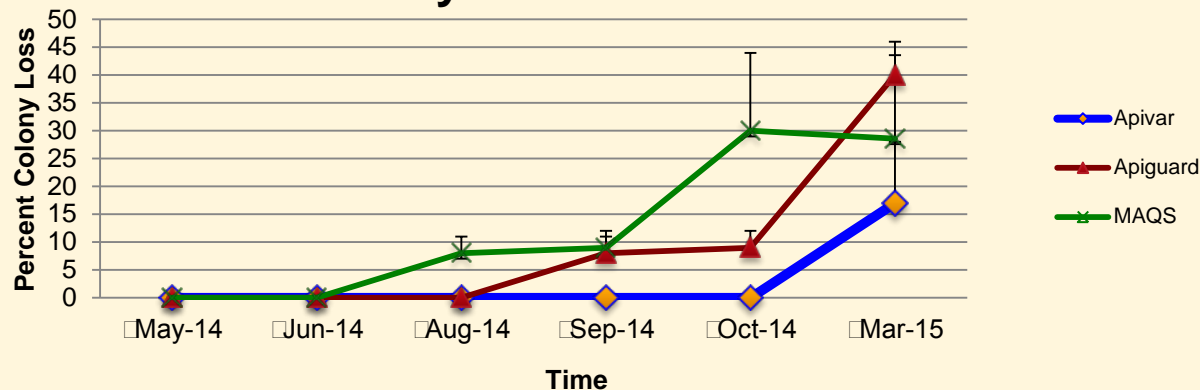
- 2014-2015:
 - Determined mite treatment efficacy
 - Treatment effect on colony strength
 - Colony losses
- 2016:
 - Install and establish bee packages
 - Implement a mite treatment regime
 - Design an IPM program



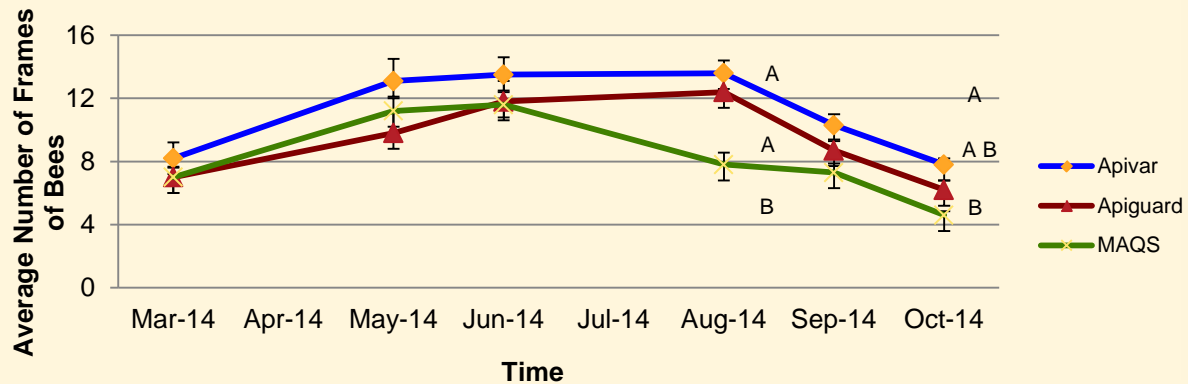
Average Fold Increase in Mites Levels Over Time



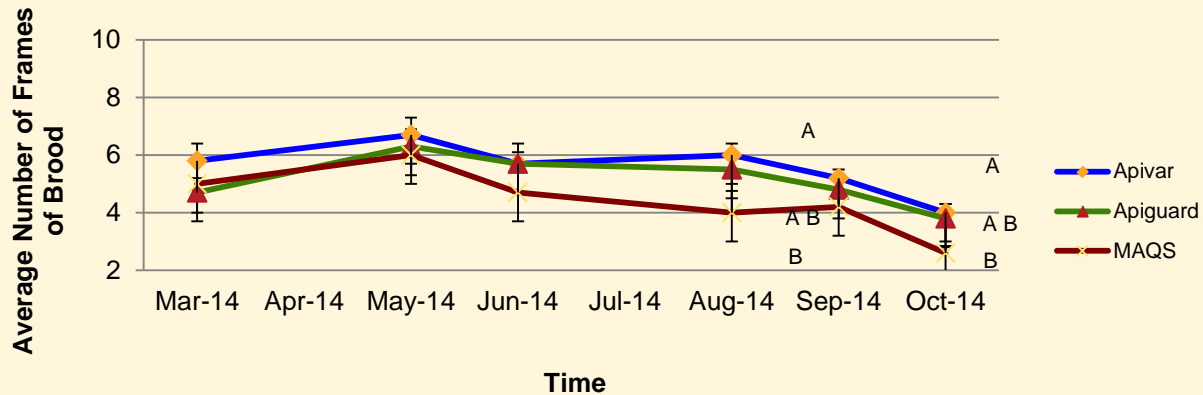
Colony Losses Over Time



Frames of Bees Over Time



Frames of Brood Over Time



Next Phase

- Install and establish bee packages
- Varroa levels baseline
- Apply Spring, Summer and Fall mite treatments
- Monitor mite levels and colony strength
- Design an Integrated Pest Management Program for Varroa control



Acknowledgements

Almond Board of California

Gene Brandi's Apiaries

Carl Hayden Bee Research Center



**Troy Anderson,
Virginia Tech**



Discovery of Stilbene Chemistries for Varroa Mites

Troy D. Anderson, PhD

Department of Entomology, Virginia Tech

Pesticide Risk Characterization for Honey Bees

Exposure

Fate, Persistence, & Application

X

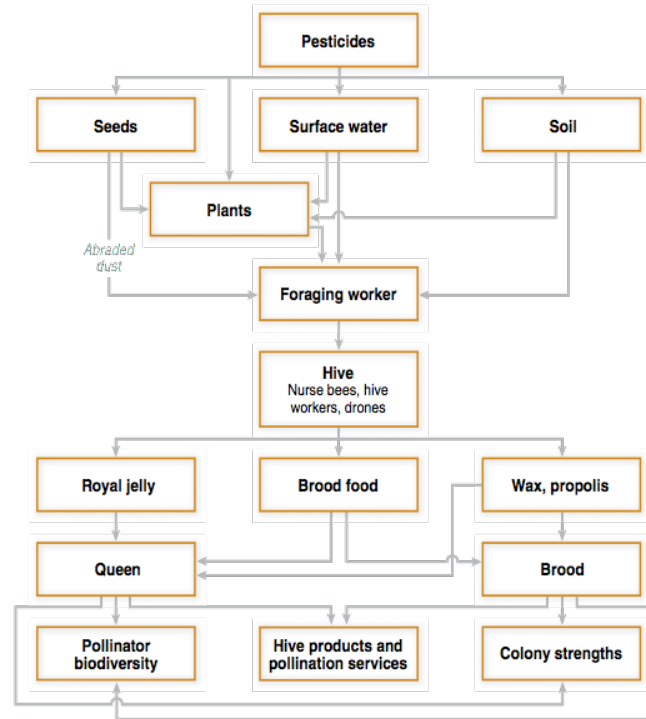
Toxicity

Laboratory vs. Field Testing

=

Risk

Predict Effects of Pesticide
Use, Misuse, & Safety



Fairbrother, Anderson et al. 2014

Honey Bee Colony Losses for Apiculture Industry

Bee Colony Losses:

~ 30% in Virginia

\$1.3-1.8 Million Lost:

Pollination Fees

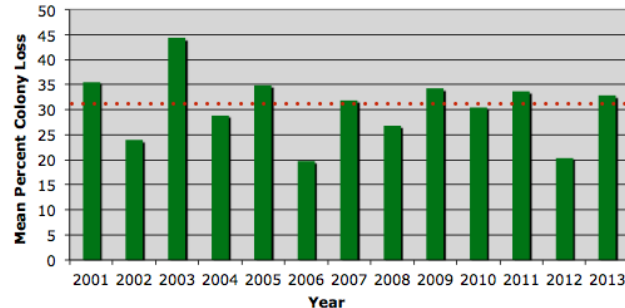
Honey Sales

Colony Replacement

Multiple Stressors,
Multiple Interactions



The screenshot shows a news article from USA Today. The title is "U.S. losing bees and beekeepers". The byline is "By Heather Collura, Special for USA TODAY". The article text states: "The number of bees is on the decline across the USA, and there's also a shortage of beekeepers. The number of commercial beekeepers is dwindling because the business of keeping bees is not as profitable as it once was, according to Jeff Pettis, research leader at the U.S. Department of Agriculture Bee Research Laboratory in Maryland. That decline in profitability is due in large part, Pettis said, to lower honey prices — the average U.S. price per pound dropped four-tenths of a cent over the past year. Keepers also face difficulty in keeping healthy bees." There is a photo of a beekeeper in a white protective suit and hat working with a hive.



Honey Bee Colony Losses for Apiculture Industry

Bee Colony Losses:

~ 30% in Virginia

\$1.3-1.8 Million Lost:

Pollination Fees

Honey Sales

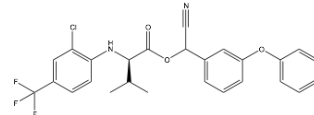
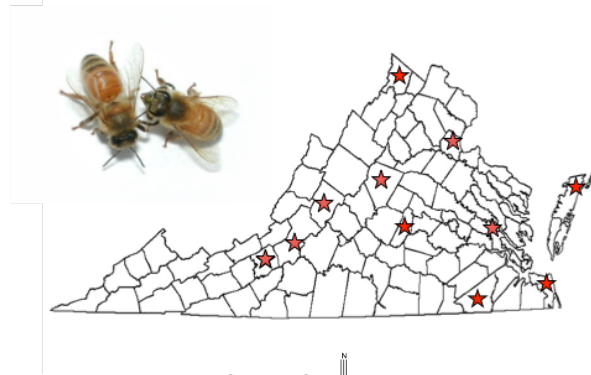
Colony Replacement

Multiple Stressors,
Multiple Interactions



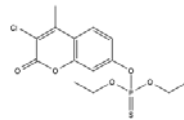
Credit: Bayer CropScience

Pesticide Residue Exposures in Honey Bees Colonies



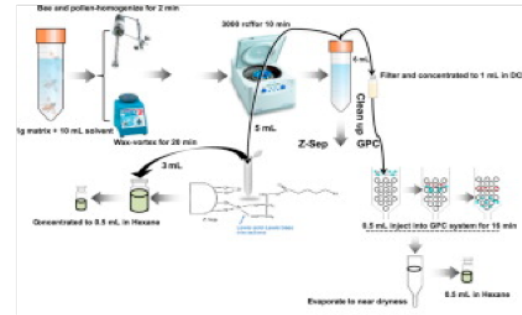
tau-Fluvalinate (LD₅₀ = 15,860 ng/g)

Bee Pollen Wax
 18 ± 1 ng/g 29 ± 2 ng/g 197 ± 11 ng/g

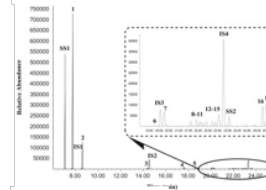


Coumaphos (LD₅₀ = 46,300 ng/g)

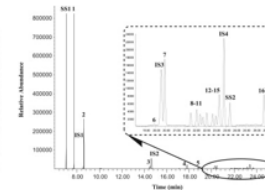
Bee Pollen Wax
 243 ± 14 ng/g 323 ± 28 ng/g 1100 ± 76 ng/g



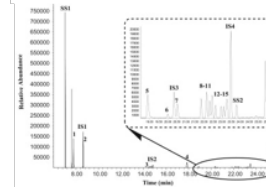
A. Surrogates



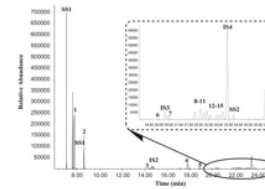
B. Bee Extracts



C. Pollen Extracts



D. Wax Extracts

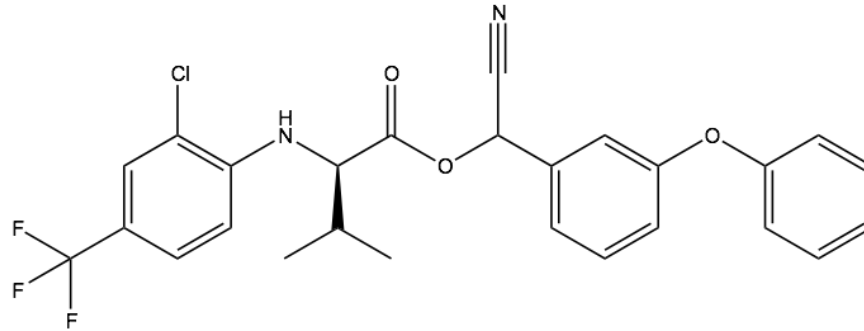


Li, Anderson et al. 2015

Major Pest Management Challenge for Apiculture Industry



In-Hive Acaricides for Varroa Mite Management



***tau*-Fluvalinate**

Cyano(3-phenoxyphenyl)methyl N-[2-chloro-4-(trifluoromethyl)phenyl]-D-valinate

Pyrethroid Class

Section 18 in 1988

VGSC Target Site

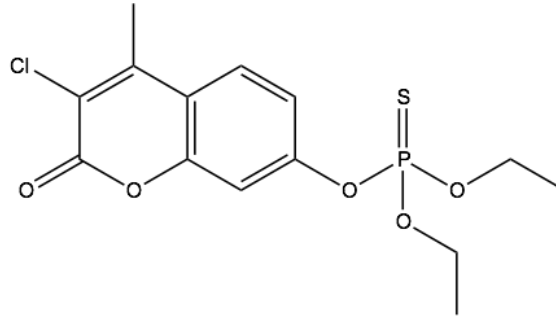
kdr Resistance

I of 4 Active
Diastereoisomers

Bee Nutrition and
Immune Health Issues

Reeves, Anderson et al. 2014

In-Hive Acaricides for Varroa Mite Management



Coumaphos

O-(3-Chloro-4-methyl-2-oxo-2H-chromen-7-yl) O,O-diethyl phosphorothioate

Organophosphate Class

Section 18 in 1999

AChE Target Site

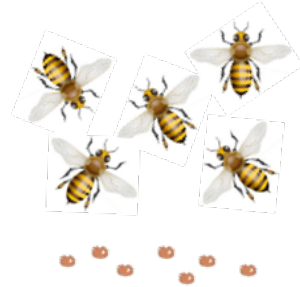
Metabolic Resistance

26+ Commercial
Products

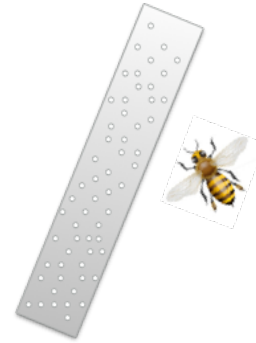
Bee Nutrition and
Immune Health Issues

Reeves, Anderson et al. 2014

In-Hive Acaricides for Varroa Mite Management



Mites are exposed to acaricide resulting in paralysis

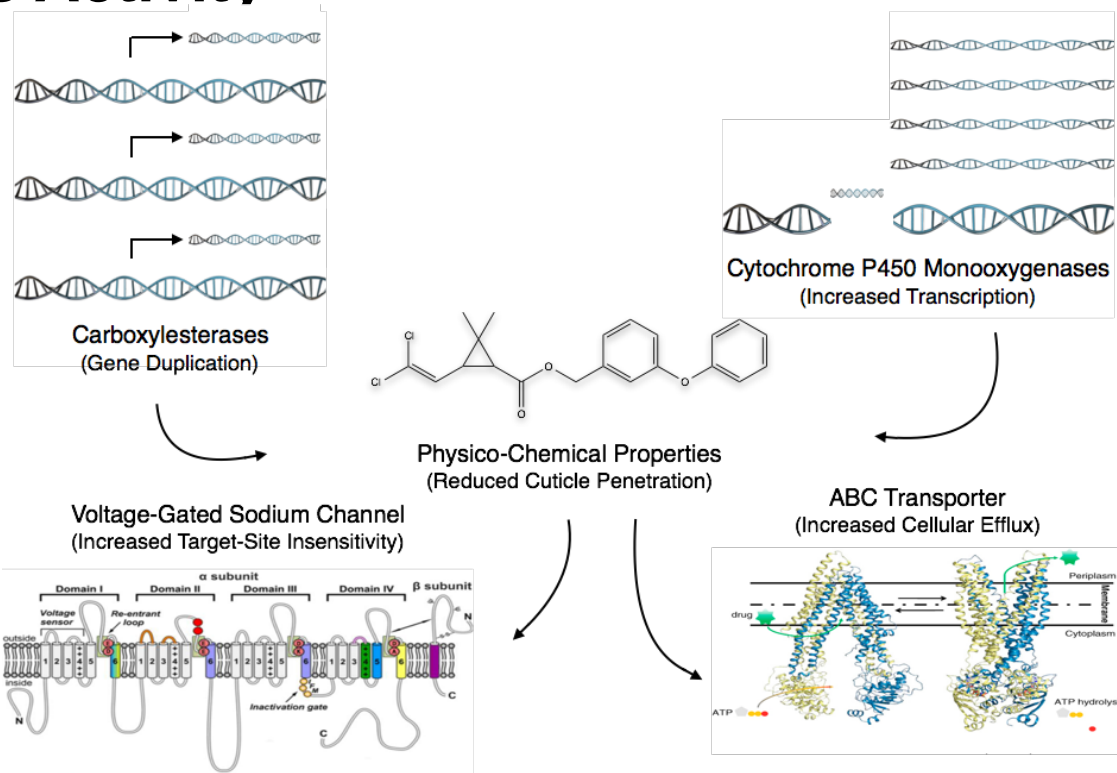


Bees walk on acaricide strips and pick up molecules



Bees distribute acaricide via contact with each other

Metabolic & Target-Site Resistance Limits In-Hive Acaricides Activity



Testing Acaricide Efficacy and Resistance in Honey Bee Colonies



Philene Vu, MS Student



Price's Fork, Kentland Farm, and Moore Farm Apiaries in Blacksburg, VA



Sample Varroa Mites from Brood Frames in Each Bee Colony



Collect ~300 Brood-Nest Bees from Each Frame for Acaricide Bioassays

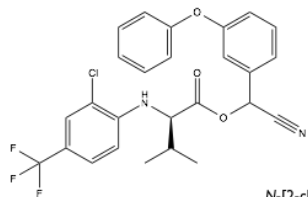


Expose Varroa Mites to Acaricide-Treated Tabs for 3- and 6-hr Intervals



Rinse Brood-Nest Bees with Ethanol to Remove Remaining Varroa Mites

Testing Acaricide Efficacy and Resistance in Honey Bee Colonies

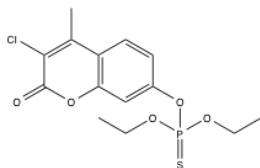


tau-Fluvalinate
(Apistan®, 10.0% ai)

Cyano(3-phenoxyphenyl)methyl
N-[2-chloro-4-(trifluoromethyl)phenyl]-D-valinate

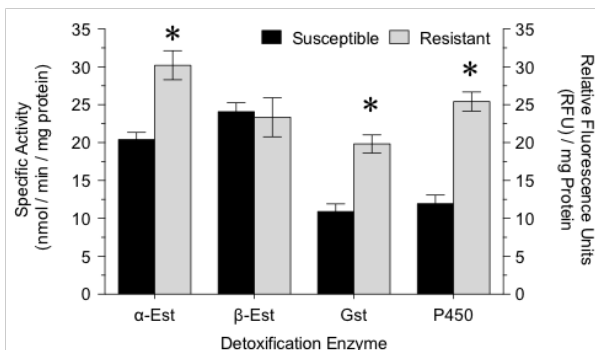
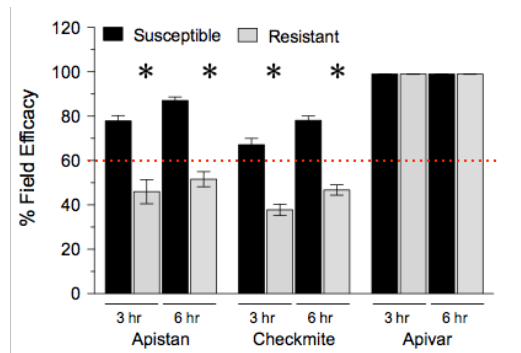
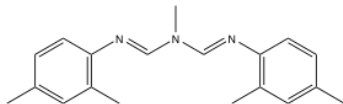
Coumaphos
(CheckMite+™, 10.3% ai)

O-(3-Chloro-4-methyl-2-oxo-2H-chromen-7-yl)
O,O-diethyl phosphorothioate



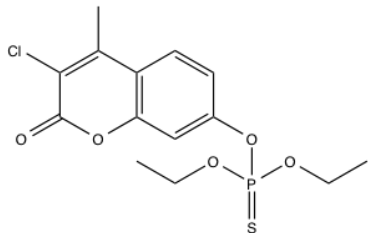
Amitraz (Apivar®), 3.3% ai)

N-(2,4-Dimethylphenyl)-N-((E)-
[2,4-dimethylphenyl]imino)methyl)-N-methylimidoforamide



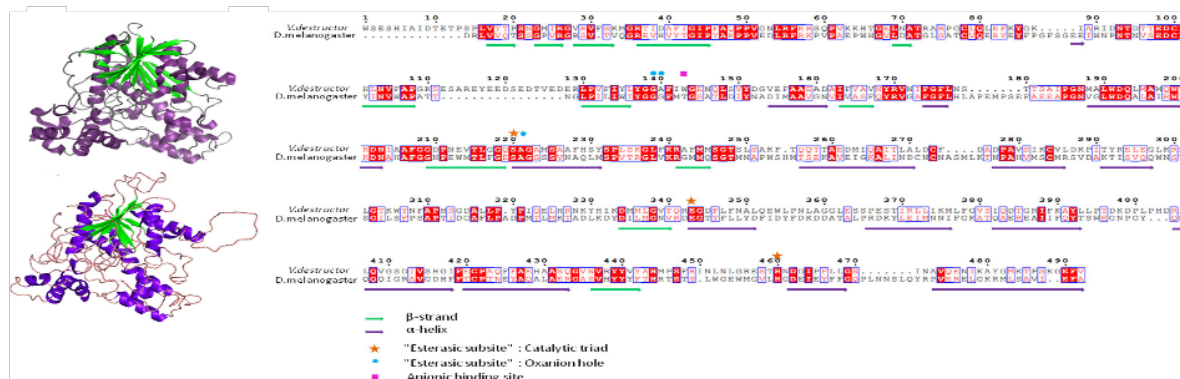
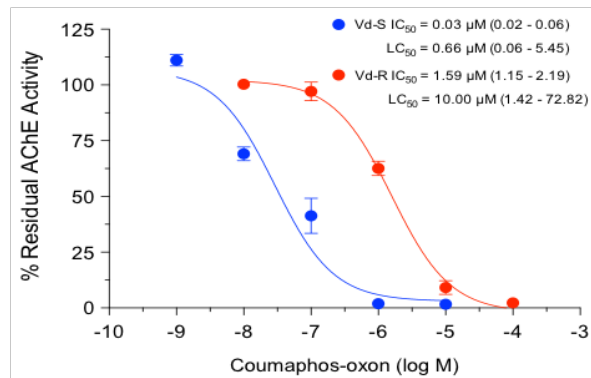
Vu, Anderson et al. 2015

Testing Acaricide Efficacy and Resistance in Honey Bee Colonies



Coumaphos
(CheckMite+™, 10.3% ai)

O-(3-Chloro-4-methyl-2-oxo-2H-chromen-7-yl)
O,O-diethyl phosphorothioate



Vu, Anderson et al. 2015

Natural Product Stilbenoid Scaffold for Alternative In-Hive Acaricides

Pest Management Science *Pest Manag Sci* 64:646–653 (2008)

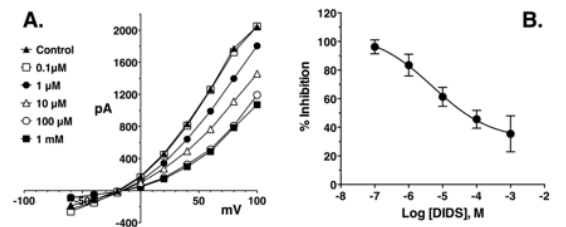
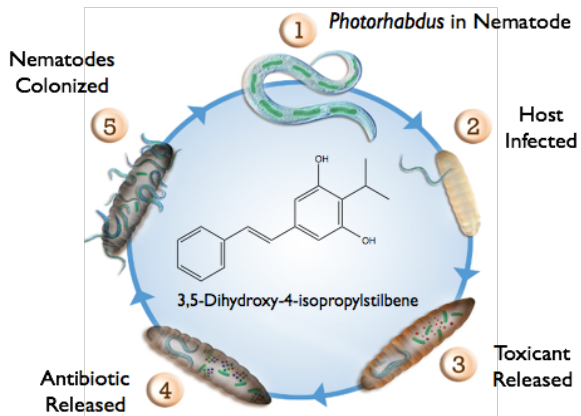
SCI
SCIENCE DIRECT

Nematicidal activity of anion transport blockers against *Meloidogyne incognita*, *Caenorhabditis elegans* and *Heterorhabditis bacteriophora*

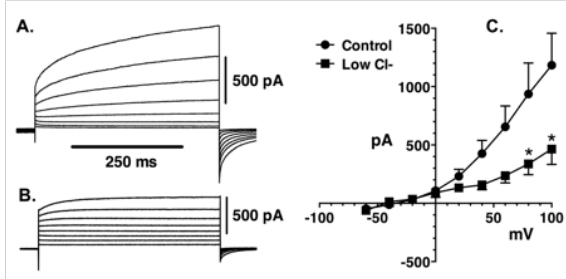
Invert Neurosci
DOI 10.1007/s10158-012-0143-8

ORIGINAL PAPER

Voltage-sensitive chloride ion channels in *Anopheles gambiae* Sua-1B cells



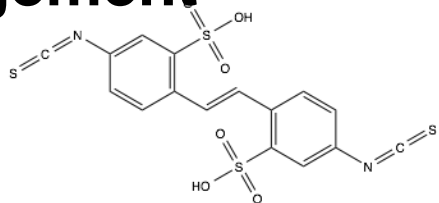
Electrophysiological effects of DIDS on chloride currents in Sua1B cells. (A) Current-voltage relationship of chloride current in Sua1B cells and inhibition with DIDS (n = 6). (B) Concentration-response curve representing percent inhibition of DIDS at +60 mV (n = 6).



DIDS insensitive chloride currents under planar patch conditions. Traces in (A) and (B) illustrate currents in normal (297 mOsmol) and low (35 mOsmol) chloride extracellular solutions, respectively. Plot (C) shows current-voltage relationship of DIDS-insensitive chloride channels in control and low chloride solutions (n = 3). Asterisks indicate a significant difference in current amplitude at a given voltage step in control or low chloride conditions (unpaired T-test, P < 0.05).

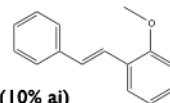
Jenson, Anderson et al. 2015, Jenson, Anderson et al. 2016

Resistance-Breaking Stilbene Chemistries for Varroa Mite Management

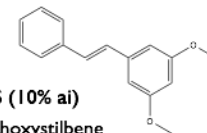


DIDS (10% ai)

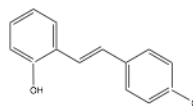
4, 4'-Diisothiocyanatostilbene-2, 2'-disulfonic acid



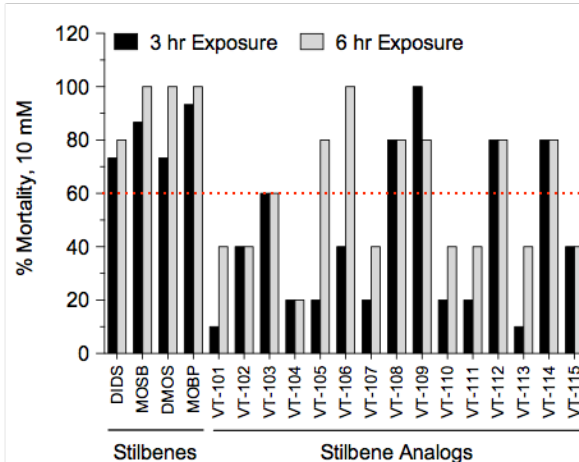
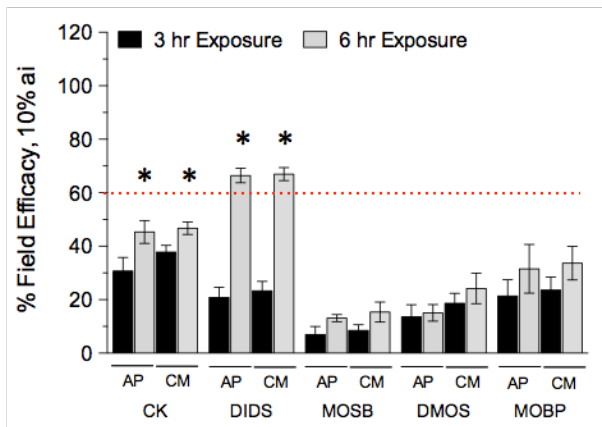
MOSB (10% ai)
2-Methoxystilbene



DMOS (10% ai)
3,5-Dimethoxystilbene



MOBP (10% ai)
(E)-2-(4-methoxystyryl)phenol



Vu, Anderson et al. 2015

Project Summary & Current Directions

Bee decline has become a nationally recognized problem, demanding attention from both the scientific community and the beekeeping industry.

Pesticide use is one of the primary perceived problems for bee decline, with *tau*-fluvalinate and coumaphos affecting the nutrition and immune health of honey bees (Reeves and Anderson 2014).

Widespread acaricide resistance limits the use of current chemistries to reduce the risk of varroa mite infestations and infectious diseases.

Stilbene chemistries provide an innovative approach for an alternative chemical strategy to deplete or incapacitate varroa mites.

Current research activities are focused on the acaricide-resistance monitoring, identification of metabolic and target-site resistance mechanisms, and discovery of alternative chemistries with acaricidal activity against varroa mites.